Title: Specific detection of *Rhizoctonia* pathogens using genome fingerprinting and hybridization based analyses

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Abstract: Several species and hyphal anastomosis groups (AG) of *Rhizoctonia solani* (sensu lato) cause brown patch diseases of turfgrasses. Conventional methods of identification of Rhizoctonia pathogens are time consuming and often inaccurate. To achieve more reliable pathogen identification, sequence characterized amplified region (SCAR) markers from Universally primed PCR (UP-PCR) products were developed to identify turf pathogenic isolates of R. solani AG 1-IB and AG 2-2IIIB. The developed SCAR markers could distinguish isolates of AG 1-IB or AG 2-2IIIB groups and did not amplify any product from genomic DNA of non-target isolates of *Rhizoctonia*. The specific primers were sensitive and unique enough to produce a PCR band from total DNA of diseased turfgrasses infected with either AG 1-IB or AG 2-2IIIB. Moreover, a rapid identification assay for Waitea circinata (anamorph: Rhizoctonia spp.) varieties zeae and circinata which cause patch diseases, was developed based on the UP-PCR products cross-blot hybridization. Isolates within a W. circinata variety crosshybridized strongly while non-homologous isolates did not cross-hybridize or did so weakly. Closely related W. circinata varieties zeae and circinata were clearly distinguished using this assay. For phylogenetic differentiation, seventy-nine previously characterized R. solani (n = 55) and W. circinata (n = 24) isolates were analyzed with AFLP markers generated by four primer pairs. Separate analysis of R. solani and W. circinata isolates based on unweighted pair group method with arithmetic mean (UPGMA) correctly grouped them according to their AG, AG subgroup, or W. circinata variety. Principle component analysis (PCA) corroborated UPGMA clusters. This is the first time AFLP analysis has been tested as a method to decipher the AG, AG subgroup, or W. circinata variety across a wide range of Rhizoctonia isolates.

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